Associations between KRAS status and clinical features in non-polyp colon cancer patients

Cuong Hoang Minh¹, Huong Bui Thi Thu², Loi Nguyen Thuan³, Thang Vu Hong¹

Abstract

Background Mutations of the KRAS gene are common in cancers; they are detected at a very high rate in colon cancer (CC) and are considered a negative disease factor. CC is among the deadliest cancers in Vietnam, and KRAS status for non-polyp colon cancer (NPCC) was rarely reported from the country. In this study, we aim to determine KRAS mutations and the associations of mutation status with clinicopathological features in Vietnamese NPCC patients.

Methods Patients with NPCC (stages II or III) and without historical or current polyp appearance were included. Genomic DNA samples were prepared from dissected tumors, and specific sequences of the KRAS gene were amplified by PCR. The mutations at codons 12, 13, 59, 60, 61, 117, and 146 of the gene were determined by using a commercial kit. Possible associations of the KRAS mutation with clinicopathological properties were analyzed using SPSS and GraphPad Prism.

Results The KRAS mutation rate was 47.9% in NPCC patients; mutations in exon 2 accounted for 91.4% of all detected mutations. Moreover, the KRAS mutation rate was higher in females (57.1%) than in males (39.8%). The association of KRAS mutation with female NPCC patients was further confirmed by multivariate regression data with OR=2.144 and p = 0.012.

Conclusion The KRAS mutation rate was also higher in patients with right colon cancers. The mutated-KRAS-carrying patients potentially experienced anemia. The data provide important scientific background for the treatment and management of the disease.

Keywords non-polyp colon cancer, KRAS mutation in NPCC, non-polyp colon cancer, stage II-III
Introduction

Colon cancer (CC) is among the five most common cancers in the world. In 2020, 1,148,515 new cases were reported and 576,858 deaths were attributable to CC worldwide [1]. Many years of biomarker discovery attempts have provided a chance of early diagnosis and partly supported mortality reduction in the CC [2]. Among important discovered oncogenes, KRAS (KRAS proto-oncogene, a homolog of Kirsten ras viral oncogene in humans) has been showing the highest detected mutation rate in CC, with over 35% of patients [3-5]. The KRAS mutations dominantly occurred in exon 2 at codons 12 and 13 [6]. Mutations at codons 59 and 61 in exon 3 were reported at low rates [7-9], while mutations at two codons of exon 4 (117 and 146) were rarely detected [10, 11]. Most of the gene mutations continuously trigger MAP-Kinase pathways and, subsequently, the proliferation of cancer cells [12, 13]. The mutated gene, therefore, is considered the negative factor [5, 14, 15], with poor disease-free survival in CC [6, 16]. Screening for mutations in this gene benefits patients when using drugs currently available and/or under development [17, 18].

CC is the fourth deadliest cancer in Vietnam (both genders combined) [19], with an increasing trend of new cases [20]. Most Vietnamese patients were diagnosed in stages II or III (by AJCC 8th) since the tumors invaded muscle layers or migrated to local nodes [16]. However, only a few studies on KRAS mutations in Vietnamese colorectal cancer, with most of the recruited patients carrying the CC, have been reported [21, 22]. The KRAS status of only non-polyp colon cancer (NPCC) has not been elucidated. Furthermore, the association of the KRAS status with the clinical and clinicopathological properties of the Vietnamese NPCC is also under establishment. Therefore, this study aims to determine KRAS mutations and their associations with clinicopathological features in Vietnamese patients who were hospitalized for surgery to remove NPCC.

Methods

Patients

Data in this study covered all patients with non-polyp colon cancer stages II or III who were hospitalized from January 2016 to August 2020 at Bach Mai, Viet Duc, and the Vietnam National Cancer Hospital—the three biggest national hospitals in northern Vietnam. Patients with colon cancer and without tumors in any other tissues or organs are included. The stage of colon cancer (II or III) was followed by the standards of the American Joint Committee on Cancer (AJCC), version 8. The term "non-polyp" described patients without a historical or current polyp appearance. Non-polyp status was examined by colonoscopy and/or confirmed by observing the dissected tissues after radical surgery. Patients with the appearance of colon polyps in any number at any time were excluded.

The Ethics Committee of Hanoi Medical University approved the study. The patients consented to participate in the study. Collecting and using personal information and clinicopathological characteristics was performed with the permission of the patients. The handling of the collected information, which was classified and safely stored as confidential, was strictly limited to assigned members.

DNA isolation and Mutation analysis

Investigation of KRAS mutations was performed with formalin-fixed, paraffin-embedded (FFPE) tissue. Five 10-m-thick tissue slides from each FFPE tissue block (194 samples) were used for the isolation of genomic DNA. Paraffin in FFPE tissue slides was removed by using FFPE deparaffinization solution (MERCK, Germany), and DNA was then extracted from tissue samples by using the PureLinkTM Genomic DNA Mini Kit (Invitrogen, USA).

KRAS mutations were detected by using the KRAS XL StripAssay (ViennaLab, Austria) in a five-step procedure including amplification, hybridization, stringent wash, and color development. Briefly, prepared DNA samples were mixed with Taq polymerase and an amplification mixture from the manufacturer. The specific DNA sequences were amplified in 35 cycles of polymerase chain reactions (PCR). The PCR products were then denatured and hybridized with probes on strips in the hybridization
buffer. After hybridization ended, solutions were removed and the strips were washed. A conjugated solution was then added to the strips, followed by a second washing step. At last, color developer was added to the strips in the dark for the appearance of purple positive bands, and then a third washing step took place. The strips were analyzed to determine KRAS mutations at codons 12, 13, 59, 60, 61, 117, and 146.

Statistics

The KRAS XL StripAssay (ViennaLab, Austria) used a five-step procedure that included amplification, hybridization, stringent wash, and color development to find KRAS mutations. Briefly, prepared DNA samples were mixed with Taq polymerase and an amplification mixture from the manufacturer. The specific DNA sequences were amplified in 35 cycles of polymerase chain reactions (PCR). The PCR products were then denatured and hybridized with probes on strips in the hybridization buffer. After hybridization ended, solutions were removed and the strips were washed. The strips then received a conjugated solution, and a second washing process followed. At last, color developer was added to the strips in the dark for the appearance of purple positive bands, and then a third washing step took place. The strips were analyzed to determine KRAS mutations at codons 12, 13, 59, 60, 61, 117, and 146.

Results

Figure 2. Positions of KRAS mutation in dissected tumors from Vietnamese patients with NPCC.

Figure 3. Clinical properties of patients with NPCC before surgery based on KRAS status.
Included patients

During the course of the study, a total of 194 patients who satisfied the selection criteria were included in the analysis. Among them, 91 (46.9%) were females, of whom 39 (20.1%) carried the wild-type and 52 (26.8%) the mutated KRAS. On the other hand, 103 (53.1%) were males, including 62 (32.0%) wild-type and 41 (21.1%) mutated KRAS. The age of the patients ranged from 24 to 75, with an average of 55.4 ± 12.2. Summarized in Figure 1, the minimum ages of the mutated KRAS-carrying males and females were lower than those of the patients with the wild-type KRAS. The average age of the mutated patients was 53.91 (54.51 for the mutated KRAS males; 53.44 for females), which is also lower than 56.77 for the wild-type KRAS group (57.37 for males; 55.82 for females) (Figure 1). However, pair-wise comparisons of the average ages showed no significant changes in age between the mutated and the normal groups (data not shown).

Mutations in KRAS gene in Vietnamese NPCC stages II and III

Point mutations of KRAS in exons 2 (codons 12 and 13), 3 (codons 59 and 61), and 4 (codons 117 and 146) were determined. The overall KRAS mutation rate was 47.9% in patients (Figure 2). Of which, mutations in exon 2 were absolutely dominant, with 85 cases accounting for 43.8% of recruited patients (or 91.4% of total detected mutations). Mutations in exon 3, including 1 case at codon 59 and 5 cases at codon 61, resulted in 3.1%. Only two cases of mutated exon 4 were detected (1%); one carried a mutation at codon 117 and the other at codon 146. None of the patients were found to carry more than one mutation (Figure 2).

Associations of KRAS status and clinical features

A pairwise comparison between the wild-type and mutated KRAS groups for each symptom was brought about to identify a possible association of a symptom with the KRAS status. Most of the observed symptoms were unlikely to correlate with the KRAS mutations (Figure 3). Abdominal pains were highly recorded in both groups, with 84.2% and 77.4% in the wide-type group and mutation groups, respectively. Other symptoms that neither showed considerable changes between the two groups included bleeding (14.9% vs. 22.6%), blood in stool (21.8% vs. 11.8%), diarrhea (25.7% vs. 26.9%), constipation (19.8% vs. 17.2%), and weight loss (19.8% vs. 20.4%). However, the analysis revealed a significant increase in anemia cases among the mutated group (p = 0.041). Particularly, 28.0% of mutated-KRAS patients exhibited anemia symptoms, while only 15.8% of the wide-type group did, indicating an association of the anemia with the KRAS mutation.

### Table 1. Clinicopathological features and KRAS status of tumor-dissected patients.

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>Total (n=194)</th>
<th>KRAS mutation status</th>
<th>P value</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Wild type N=(101, 52.1%)</td>
<td>Mutated (n=93, 47.9%)</td>
<td></td>
</tr>
<tr>
<td>Age &lt; 50</td>
<td>56</td>
<td>28(27.7%)</td>
<td>28(30.1%)</td>
</tr>
<tr>
<td>≥50</td>
<td>138</td>
<td>73(72.3%)</td>
<td>65(69.9%)</td>
</tr>
<tr>
<td>Gender Male</td>
<td>103</td>
<td>62(61.4%)</td>
<td>41(44.1%)</td>
</tr>
<tr>
<td>Female</td>
<td>91</td>
<td>39(38.6%)</td>
<td>52(55.9%)</td>
</tr>
<tr>
<td>Tumor location Right colon</td>
<td>92</td>
<td>41(40.6%)</td>
<td>51(54.8%)</td>
</tr>
<tr>
<td>Left colon</td>
<td>102</td>
<td>60(59.4%)</td>
<td>42(45.2%)</td>
</tr>
<tr>
<td>Histology AC</td>
<td>164</td>
<td>84(83.2%)</td>
<td>80(86.0%)</td>
</tr>
<tr>
<td>Others</td>
<td>30</td>
<td>17(16.8%)</td>
<td>13(14.0%)</td>
</tr>
<tr>
<td>T3</td>
<td>81</td>
<td>47(46.5%)</td>
<td>34(36.6%)</td>
</tr>
<tr>
<td>Invasion depth T4a</td>
<td>92</td>
<td>43(42.6%)</td>
<td>49(52.7%)</td>
</tr>
<tr>
<td>T4b</td>
<td>21</td>
<td>11(10.9%)</td>
<td>10(10.8%)</td>
</tr>
<tr>
<td>N0</td>
<td>93</td>
<td>52(51.5%)</td>
<td>41(44.1%)</td>
</tr>
<tr>
<td>N1</td>
<td>71</td>
<td>32(30.7%)</td>
<td>40(43.0%)</td>
</tr>
<tr>
<td>N2</td>
<td>30</td>
<td>18(17.8%)</td>
<td>12(12.9%)</td>
</tr>
<tr>
<td>II</td>
<td>93</td>
<td>52(51.5%)</td>
<td>41(44.1%)</td>
</tr>
<tr>
<td>III</td>
<td>101</td>
<td>49(48.5%)</td>
<td>52(55.9%)</td>
</tr>
</tbody>
</table>

AC: adenocarcinoma; *: There were statistical significance.
Table 2. Interactions between KRAS mutation and prognostic markers.

<table>
<thead>
<tr>
<th>KRAS mutation</th>
<th>Multivariate logistic regression</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>OR</td>
</tr>
<tr>
<td>Age</td>
<td>≥50 / &lt; 50</td>
</tr>
<tr>
<td>Gender</td>
<td>Femal / Male</td>
</tr>
<tr>
<td>Tumor location</td>
<td>Left / Right</td>
</tr>
<tr>
<td>Invasion depth</td>
<td>T4 / T3</td>
</tr>
<tr>
<td>Nodal status</td>
<td>N(+) / (-)</td>
</tr>
<tr>
<td>Histology</td>
<td>AC / Others</td>
</tr>
</tbody>
</table>

OR: odds ratio; AC: Adenocarcinoma.

in NPCC (Figure 3).

Association of KRAS mutation and clinicopathological features in patients with NPCC stages II and III

Table 1 demonstrated possible associations of clinicopathological features with KRAS status in patients. Conspicuously, the KRAS mutation rate in females was statistically higher than in males. With 52 mutated females, the KRAS mutation rate was 55.9% of the cases, far higher than the 44.1% contributed by the males (Table 1). The mutations were also detected at a higher rate in tumors in the right colon compared with the left colon (54.8% right colons vs. 45.2% left colons, p = 0.047) (Table 1). In contrast, KRAS mutation rates were relatively compatible among younger and older patients (Table 1).

Histologic examination showed a high rate of adenocarcinoma (AC) with 164 confirmed cases (84.5% of patients), while other types were identified in 30 cases (15.5% of patients) (Table 1). The numbers of NPCC AC cases in mutated vs. wild-type KRAS were almost identical, 80 and 84, respectively, indicating that the KRAS mutation is unlikely to cause the AC. The KRAS mutation rate among the AC was also not different from that among the other types.

The invasion depth of tumors in Vietnamese NPCC was likely linked to the KRAS mutation. The T4 stage (which included T4a and b) was confirmed in 59 cases of the mutated group, accounting for 63.5%. This stage was confirmed in only 54 wild-type patients (53.5% of the wild-type group). Similarly, stage III tumors were determined at a higher rate among the mutated patients (52 out of 93 cases, or 55.9%) compared with the same stage among the wild-type patients (49 out of 101 cases, or 48.5%). However, none of the linkages between the staging and KRAS mutation were statistically significant, including nodal stages (Table 1).

Multivariate logistic regression analysis: interaction of KRAS mutations with other prognostic markers

The association of KRAS mutations with female among Vietnamese NPCC patients was confirmed by multivariate logistic regression. The KRAS mutation rate was significantly higher in females, with OR = 2.144 (95% confidence interval (CI): 1.184 – 3.882, p = 0.012). The analysis further confirmed the correlation of KRAS mutations with the right NPCC, with p = 0.048. Therein, the odds ratio of left colon tumors to right colon tumors was 0.551 (95% confidence interval (CI): 0.305-0.996) (Table 2). Besides the differences above, there was no evidence of an association between tumor staging and histologic types and KRAS mutations among NPCC in Vietnam.

Discussion

The association of mutations in the KRAS gene with poor disease-free survival in colon cancer has been well studied worldwide [6]. However, the KRAS mutation rate in CC, especially in NPCC, was rarely reported from Vietnam. In this study, we found a notable high KRAS mutation rate (47.9%) among patients with NPCC in the country. The detected rate was surprisingly higher than that in previous reports for CC, which showed KRAS mutation rates around 36-38% [3, 23, 24]. The change mostly occurred due to the mutation rate in exon 2 of the gene. Particularly, the mutation rate in KRAS exon 2 was 43.8% of the recruited patients, dramatically higher than that in CRC in Vietnam (37.1%) [22] and China (42.2%) [25]. The mutation rate in KRAS exon 3 (3.1%) was relatively similar to that in a previous study [25], while the mutations in exon 4 were rare in Vietnamese NPCC (only 2 cases, or 1%).

Another impressive finding in this study was the association of KRAS mutations with female patients. The mutations were detected in 57.1% of female patients (52 mutants out of 91 females) and only 39.8% of male patients (41 mutants out of 103 males). Females accounted for 55.9% of mutated patients (Table 1), while males were 44.1%. These gender ratios among mutated patients were contrary to those in a previous study for CRC in Vietnam, which reported 48.2% and 51.8% for mutated females and males, respectively [22]. The contrary suggested an association of KRAS mutations with the NPCC in females, which was further confirmed by our multivariate logistic regression data shown in Table 2. Besides, the age of KRAS mutation-carrying patients in both genders was apparently lower than that of the wild-type groups. However, the correlation of the KRAS mutation with the patient’s age was unclear.

Several reports showed that the KRAS mutation rate was likely higher in the right colon than in the left [3, 18, 26]. In the present study, the linkage of the KRAS mutation rate with right colon tumors was clarified. The KRAS mutation rate in the right colon was 55.4% (51 mutants out of 92), while the rate in the left colon was only 41.2% (42 mutants out of 102). Multivariate regression
analysis showed significantly higher KRAS mutations in right colon tumors with $0.048$. The right colon cancers, in comparison with the left ones, were previously reported to be linked with worse survival of the patients [3]. A further study is required to investigate the survival of Vietnamese patients with NPCC.

Similarly, clinicopathological properties such as abdominal pain, anemia, and various symptoms were identified in over 83% of patients. Of those, diarrhea and anemia were likely correlated (close to statistically significant change) to the tumors in the right colon [27]. Logically, a linkage between diarrhea and anemia with KRAS mutations is suspected. However, our current data has confirmed significantly higher anemia cases but not diarrhea cases among KRAS-mutated NPCC patients.

**Conclusion**

The KRAS mutation rate was very high (47.9%) in Vietnamese NPCC patients. The mutation in exon 2 was absolutely dominant, accounting for 91.4% of detected mutations. Moreover, the KRAS mutations likely occurred in Vietnamese females with NPCC (OR=2.144, $p = 0.012$), as well as in right colon cancer patients. The patients who carried the mutated KRAS further potentially experienced anemia before being diagnosed with the NPCC. These findings provide important scientific background for the treatment and management of the NPCC.

**Acknowledgements**

None.

**Availability of data and materials**

Data and materials are available on request from the authors.

**Ethical policy**

The study was approved by the Ethics Committee of Hanoi Medical University (Approval number: NCS28/HMU-IRB).

**Author contributions**

MCH conceptualized, designed, conducted research, and wrote the first draft. TDL, TTHB, TLN, TJA, and HTV provided supervision and revision of the draft.

**Competing interests**

The authors have no conflicts of interest regarding the publication of this paper.

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